

THE STRUCTURE OF EPILUBIMIN, A STRESS METABOLITE FROM
DISEASED POTATO TUBERS¹⁾

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The structure of epilubimin, a stress metabolite isolated from diseased potato tubers, was determined as shown in formula λ on the basis of the chemical and spectral data.

In a continuing study on phytoalexins produced by tuber tissues of white potatoes (*Solanum tuberosum* and *S. demissum*) infected by an incompatible race of *Phytophthora infestans*,²⁾ we isolated a new sesquiterpene, designated as epilubimin, in a $2 \times 10^{-5}\%$ yield. The present paper describes the isolation and structure elucidation of the stress metabolite.

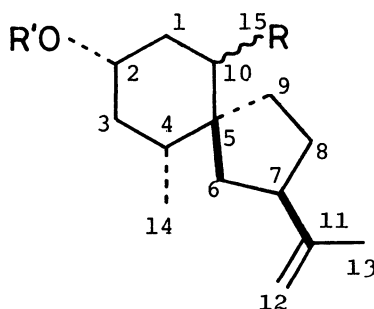
Neutral ether extracts (51 g), obtained from the diseased potato tubers (300 kg), were separated by column chromatography (CC) over silica gel (Merck Kieselgel 70-230 mesh) to give a "lubimin and epilubimin fraction"³⁾ (1.7 g) as eluates with benzene and ether (5:1 and 3:1), which was further purified by repeated CC followed by preparative TLC⁴⁾ over silica gel (Merck GF-254) to yield lubimin^{2a)} (λ , 360 mg) (more polar) and epilubimin (λ , 59 mg) (less polar) in pure state.

Epilubimin (λ), colorless oil, $[\alpha]_D^{20}$,⁵⁾ $C_{15}H_{24}O_2$, gave its monoacetate (λ_a), oil, and a glycol (λ), mp 135-136°C, on hydride reduction ($NaBH_4$), which also formed its diacetate (λ_a), oil. The MS, IR and NMR spectra⁵⁾ indicated that λ contains the following structural units: CH_3CH- [λ , δ 0.94 (3H, d J = 6.5 Hz); λ , δ 0.85 (3H, d J = 7 Hz)]; $CH_2=C(CH_3)-$ [λ , ν_{max} 1652 and 888 cm^{-1} , δ 1.72 (3H, s) and 4.68 (2H, s)]; λ , ν_{max} 1640 and 891 cm^{-1} , δ 1.68 (3H, s) and 4.64 (2H, s)]; -CHO [λ , ν_{max} 1715 cm^{-1} , δ 9.81 (1H, s)]; λ_a , no absorption near 3400 cm^{-1} and near δ 9]; -CH(OH)- [λ , ν_{max} 3300 cm^{-1} , δ 3.69 (1H, br W_H = 25 Hz); λ_a , ν_{max} 1740 cm^{-1} , δ 4.64 (1H, br W_H = 25 Hz)]. The ¹³C NMR spectra (Table 1) of λ and λ_a revealed the presence of a quaternary carbon atom (■) of spiro type (λ , δ 46.2; λ_a , δ 46.2) in λ .⁶⁾ Addition of 0.25 mol equiv of the shift reagent Eu(dpm)₃ to the chloroform-d solution of λ effected down-field shift of the NMR signals, leading to separation of most of the protons (Fig. 1), and spin-decoupling studies on the spectrum elucidated the existence of a structural moiety: (■?)-(CH₃)CH-CH₂-CH(OH)-CH₂-CH(CHO?)-(■). All the results suggested that epilubimin (λ) would be an epimer (with an axial formyl group) of lubimin (λ) (with an equatorial), though only a sharp singlet due to the formyl proton was observed in each of the NMR spectra of λ and λ_a irrespective of the presence of the shift reagent. Compound λ was then treated with base (4% KOH in

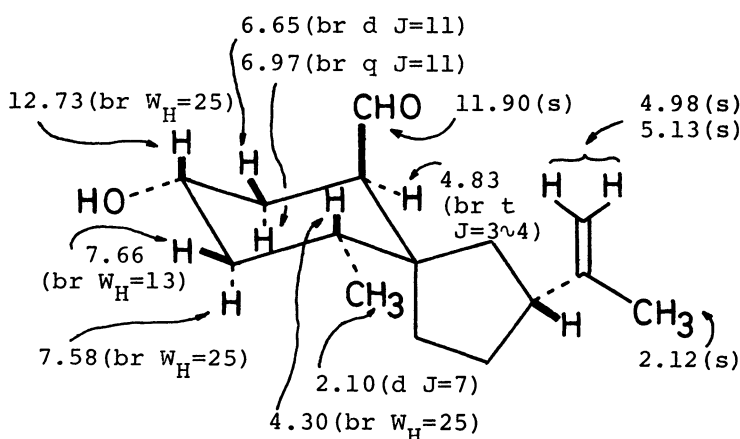
CH₃OH, room temp, 1 h) to give a mixture, from which lubimin, oil, [α]_D +31°, identical with natural lubimin (λ), oil, [α]_D +36°, in the MS, IR and NMR spectra, was isolated in a 60% yield. Hence epilubimin must be represented correctly by formula λ .

Table 1. The ¹³C NMR spectra of λ and λ_a (25.2 MHz, CDCl₃, TMS)

Compd	λ	λ_a
Carbon	Chemical shift (δ)	Chemical shift (δ) a)
1	48.9	41.3
2	66.0	69.1
3	42.1	36.0
4	35.2	35.2 or 44.0
5	46.2	46.2
6	31.1	31.1
7	47.8	47.5
8	40.0	31.5
9	31.1	30.4
10	35.2	44.0 or 35.2
11	147.9	147.3
12	107.8	108.0
13	21.3	20.9
14	17.1	16.8
15	61.2	63.9



λ R=β-CHO, R'=H
 λ_a R=β-CHO, R'=Ac
 λ R=α-CHO, R'=H
 λ R=β-CH₂OH, R'=H
 λ_a R=β-CH₂OAc, R'=Ac



a) Cf., ref. 6.

Fig. 1. The NMR spectrum (100 MHz, CDCl₃) of λ in the presence of the shift reagent Eu(dpm)₃.

REFERENCES and FOOTNOTES

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- a) N. Katsui, A. Matsunaga, H. Kitahara, F. Yagihashi, A. Murai, T. Masamune, and N. Sato, Bull. Chem. Soc. Jpn., 50, 1217 (1977), and their previous papers.
b) For a recent review, see, A. Stoessl, J. B. Stothers, and E. W. B. Ward, Phytochemistry, 15, 855 (1976).
- The fraction corresponds to "fraction E" in "Isolation of Rishitin;" T. Masamune, A. Murai, M. Takasugi, A. Matsunaga, N. Katsui, N. Sato, and K. Tomiyama, Bull. Chem. Soc. Jpn., 50, 1201 (1977).
- Cf., ref. 2a, "Isolation of Lubimin, Oxylubimin, and Lubiminol" (p. 1222).
- All the compounds described herein gave MS, IR and NMR spectra in good accord with the assigned structures. The optical rotations, IR and NMR spectra were measured in ethanol, in liquid (oil) or chloroform (λ), and in chloroform-d, respectively.
- Cf., ref. 2a, Table 1 (p. 1218).

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